

Relative Gene Expression of RND-Type Efflux Pumps in Tigecycline Resistant *Acinetobacter Baumannii* Isolated from Training Hospitals in Tehran, Iran

Mina Owrang¹, Abdollah Karimi², Leila Azimi², Reihane Motaghi Nezhad³, ^{*}Fatemeh Fallah²

¹Faculty of Medicine, Sari branch, Islamic Azad University, Sari, Iran.

²Pediatric Infections Research Center, Research Institute of children's Health, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

³Department of Microbiology, Farahan Branch, Islamic Azad University, Arak, Iran.

Abstract

Background: Appearance of multi-drug resistance (MDR) Acinetobacter baumannii imposes limitation on antibiotic therapy in patients. Detection of MDR A. baumannii can play a crucial role to prevent MDR strains spreading in hospitals. The aim of this study was determination the efflux pumps gene expression in tigecyclin resistance strains in collected isolates from selected training hospitals in Tehran, Iran.

Materials and Methods: In this cross sectional study, *A. baumannii* was collected from July to February 2014. Tigecycline susceptibility testing has been prepared according to CLSI guide- lines after identification. Active efflux pumps have been detected by Carbonyl cyanide m-chlorophenyl hydrazone (CCCP) as an efflux pumps inhibitor. Gene expressions of these efflux pumps have been determined by Real- Time PCR.

Results: In this study 80 *A. baumannii* have been confirmed by conventional phenotypic methods. Tigecyclin resistant was confirmed according to antibiotic susceptibility testing results. The results of CCCP indicated that 22.5% of tigecycline resistant *A. baumannii* could include active efflux pumps. The results of Real- Time PCR indicated that *abeM* gene expression has been observed in the most of CCCP positive *A. baumannii* and *adeB* has been observed in the minimum number of strains.

Conclusion

According to the results of this study, Efflux pumps can play an important role in appearance of cross resistance and make MDR strains. Thus, the detection of antibiotic resistance related to active efflux pumps may be crucial to find a composition with efflux pump inhibitor effect by clinical usage.

Key Words: Acinetobacter baumannii, Efflux pump, Tigecycline.

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*Corresponding Author:

Prof. Fatemeh Fallah, Medical Microbiologist, Pediatric Infections Research Center, Mofid Children Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran. P.O. Box. 15468-15514, Iran.

Email: fafallah@sbmu.ac.ir

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1- INTRODUCTION

Acinetobacter baumannii is one of the important gram-negative bacteria can cause healthcare associated infections (1-3). A. baumannii can survive in different hospital environments and also acquire antibiotic resistance genes (4, 5). Thus, it can make therapeutic complication and nosocomial infection among hospitalized patients (1- 5). Multi-drug resistance strains of A. baumannii are increasing worldwide and make therapeutic problems in healthcare centers (3- 5). Tigecycline is the first representative of the glycylcycline class of antibiotic to be marketed for treatment of complicated infection such as infectious disease caused by MDR A. baumannii isolates (6, 7). In some studies tigecycline has shown excellent in vitro activity against MDR A. baumannii. strains Tigecycline resistant of Α. baumannii are rare worldwide (6, 7).

Hence, resistance to this antibiotic can be very considerable in healthcare system because of this antibiotic and also in some case with colistin can be remain a last choice for antibiotic therapy for treatment infections caused by MDR A. baumannii strains (6, 7). Efflux pumps are one of the antibiotic resistant mechanisms that can cause cross resistance in MDR A. baumannii (4, 8). RND-type efflux pump can be involved in resistance to tigecycline (9, 10). In this regards, investigation of efflux pump effect on tigecycline resistant maybe the first step to prevent this antibiotic resistant mechanism. This study aimed to determine the relative gene expression of RND-type of efflux pumps effect on tygecycline resistant in MDR A. baumannii.

2- MATERIALS AND METHODS

2-1. Bacterial strains

In this cross sectional study 80 *A*. *baumannii* isolates were collected from different clinical samples from July to

February 2014. All isolated *A. baumannii* in microbiology laboratory of selected training hospitals were included in this study and non *A. baumannii* strains during samples collection were in exclusion criteria. The specimens were collected from pediatrics and adults. Identification of collected strains has been confirmed by conventional biochemical and microbiological tests such as; oxidase, TSI and growth on 42 $^{\circ}$ C (3, 4).

2-2. Antibiotic susceptibility testing

Tigecycline susceptibility testing has been conducted by disc diffusion method and minimum inhibitory concentration (MIC) according to CLSI guidelines (11). Antibiotic disc and tigecycline powder have been prepared from MAST Company and sigma (Sigma-Aldrich, cat No. PZ0021). *Pseudomonas aeruginosa* ATCC 27853 used as a control for antibiotic susceptibility testing and MIC.

2-3. PCR detection of efflux pumps

Conventional PCR was conducted for detecting *adeB* and *abeM* genes after DNA extraction. The primer sequences and PCR conditions have described previously (12, 13). Distilled water was used as negative controls and internal positive control after sequencing was used in this study. PCR products were run on 1% agarose gel (Sigma-Alderich, France) stained with DNA safe stain (SinaClon Co., Iran) at 85 V for one hour. Finally data were obtained by using Gel document. Direct sequencing of PCR amplified products was carried out using ABI 3730X capillary sequencer (Genfanavaran, Macrogen, Seoul, Korea). A. baumannii ATCC 19606 was used as reference stains.

2-4. Phenotypic detection of efflux pumps

Carbonyl cyanide 3chlorophenylhydrazone (CCCP) (C2759 Sigma-Alderich, France) as an efflux pumps inhibitor was used for phenotypic screening of active efflux pumps. Tigecycline MIC with CCCP (25 µg/ml) (14) in comparison with tigecycline MIC without CCCP were prepared for screening of active efflux pumps. Tigecycline for MIC, with and without CCCP, has been arranged from 0.5 to 256 µg/ml. At least four fold decrease of MIC with CCCP compared tigecycline MIC without CCCP shown presence of active efflux pumps (4, 8). A. baumannii ATCC 19606 was used as references stains.

2-5. RNA Extraction

The strains were cultured in Brain Hurt Infusion (BHI) medium (5mL) (Merck, Germany) and were grown to midexponential phase (OD600 = 1.5-2.0). Then, the bacteria $(5*10^8)$ were added to 0.5 mL of RNeasy bacteria protect solution (Qiagen, 74104, Germany) to extract RNA according to the supplier's instructions. Furthermore, DNA was eliminated using 20U of RO1 RNAse-free DNAse (Promega, Madison, WI, Korea), and was suspended in 50 µL of DEPC-treated water (0.1% v/v). A. baumannii ATCC 19606 was used as reference strains.

2-6. cDNA Synthesis

The RNA sample $(1 \ \mu g)$ was incubated with 250 ng random hexamer primers

(Sigma-Alderich, France) and was added to the pre-mix cDNA synthesis kit (BioNEER, Cat. No. K- 2041. Korea). The reaction was performed for 60 seconds at $15 \degree C$, 60 min at 55 $\degree C$ and then at 95 $\degree C$ for 5 min.

2-7. Relative gene expression of efflux pumps by Real-Time PCR

Semi quantitative Real- Time PCR has been prepared to express *adeB* and *abeM* genes; 16 srRNA (14) and *A. baumannii* ATCC 19606 have been used as a housekeeping gene and reference strain, respectively. The qPCR primers recommended by other authors were used, as shown in **Table.1**.

A Rotor Gene RT-PCR machine (Corbett Research, Sydney, Australia; Model RG3000, software version 6) was used for the duplicated PCR reactions with the Quanti Test SYBR Green RT-PCR Kit (Qiagen, Cat. No. 204243, Germany). After activation of the modified Taq polymerase at 95 ° C for 12 minutes, 40 cycles of 15 sec at 95 ° C, 30 sec at each gene annealing temperature set up and 30 sec at 72 ° C were performed. Then, the $\Delta\Delta$ CT values were used for data analysis (15). A. baumannii ATCC 19606 was used as references strains.

Target	5'-3'	Refernces
adeB	Forward AACGGACGACCATCTTTGAGTATT	12
	Reverse CAGTTGTTCCATTTCACGCATT	
abeM	Forward TGCCAATTGGTTTAGCTGTG	13
	Reverse TACTTGGTGTGCGGCAATAA	
16srRNA	Forward AACGGACGACCATCTTTGAGTATT	14
	Reverse CAGTTGTTCCATTTCACGCATT	

Table.1: Sequencing of primers in this study.

3- RESULTS

In this study 80 A. baumannii isolates have been identified and confirmed by conventional phenotypic methods. According to antibiotic susceptibility testing and MIC results, 98% (78/80) of strains were resistant to tigecycline. The of CCCP indicated results that 22.5%(18/80) of tigecycline resistant A. baumannii isolates can include active efflux pumps with a minimum 4 fold decrease of MIC + CCCP in contrast with MIC of tigecycline without CCCP. Efflux pump genes have been detected in 98% of tigecycline resistant strains by PCR as a molecular method. The results of Real-Time PCR indicated an abeM gene expression increase from 16 to more than 256 times compared to ATCC strain and adeB which increased the gene expression from 2 to 8 times in comparison with ATCC strains. Fifty percent and 70% percent of strains with CCCP positive test include an increase in adeB and abeM gene expression respectively, according to Real- Time PCR results. Actually, 4 strains showed over gene expression in both two efflux pumps.

4- DISCUSSION

The appearance of MDR and / or XDR A. baumannii is an increasing problem all over the world (4. 5). Several antibiotic resistance mechanisms can lead to the existence of MDR and XDR strains such as active efflux pumps (4, 5, 8). Efflux pumps can make resistance to several antibiotics simultaneously even in different families of antibiotics and they can be one of the important causes of the appearance of cross resistance in MDR strains (4, 8-10, 16, 17). Tigecycline is used for treatment of MDR A. baumannii (9, 10), but tigecycline resistant strains have been reported in some studies (16-20). These studies show the involvement of gene expression increase in *adeB* and abeM efflux pumps for resistance to

tigecycline in A. baumannii (18-20). In this study, 18% of tigecycline resistant A. baumannii isolates included an increasing gene expression at least in one tested efflux pumps and 16% of them showed an increase of gene expression in more than two efflux pumps. In Peleg et al. and Deng et al. study on tigecycline, non-susceptible A. baumannii showed that increase of expression in *adeB* and *abeM* in tigecycline non-susceptible isolates (18, 19). Yuhan et al. showed linear relationship between tigecycline MIC and adeB expression (20). Also, the results of this study indicated that increase in *adeB* in 50% of tigecycline resistant strains that have CCCP positive test. According to the results of this study and other studies (4, 8-10, 16- 21), efflux pumps can play an important role in resistance to tigecycline and make more complication in antibiotic therapy in A. baumannii as a one of important threats worldwide (22).

5- CONCLUSION

In this regards, the detection of *A*. *baumannii* includes active efflux pumps may be the first step to prevent the spread of the MDR strains in healthcare centers. On the other hand, the result of this study can be helpful for finding a natural substance that can inhibit efflux pumps in clinical usage. Because of inactivation of this active efflux pumps can be helpful for increasing the chance of patient's treatment especially in pediatrics.

6- CONFLICT OF INTEREST: None.

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8- REFERENCES

1. Azimi L, Motevallian A, Ebrahimzadeh Namvar A, Asghari B, Lari AR. Nosocomial infections in burned patients in motahari hospital, Tehran, Iran. Dermatol Res Pract. 2011; 2011: 436952.

2. Alaghehbandan R, Azimi L, Rastegar Lari A. Nosocomial infections among burn patients in Teheran, Iran: a decade later. Ann Burns Fire Disasters. 2012; 25: 3-7.

3. Owlia P, Azimi L, Gholami A, Asghari B, Lari AR. ESBL- and MBLmediated resistance in Acinetobacter baumannii: a global threat to burnt patients. Infez Med. 2012; 20: 182-7.

4. Ardebili A, Talebi M, Azimi L, Rastegar Lari A. Effect of Efflux Pump Carbonyl Cyanide 3-cholorophenylhydrazone on the Minimum Inhibitory concentration of ciprofloxacine in Acinetobacter baumannii Clinical Isoltes. Jundishapur J Microbiol. 2014;7:e8691

5. Azimi L, Talebi M, Pourshafie MR, Owlia P, Rastegar Lari A. Characterization of Carbapenemases in Extensively Drug Resistance Acinetobacter baumannii in a Burn CareCenter in Iran. Int J Mol Cell Med. 2015; 4: 46-53.

6. Pachón-Ibáñez ME, Jiménez-Mejías ME, Pichardo C, Llanos AC, Pachón J. Activity of Tigecycline (GAR-936) against Acinetobacter baumannii strains, including those resistant to imipenem. Antimicrob Agents Chemother. 2004; 48: 4479-81.

7. Bosó-Ribelles V, Romá-Sánchez E, Carmena J, Cáceres C, Bautista D. Tigecycline: a new treatment choice against Acinetobacter baumannii. Recent Pat Antiinfect Drug Discov. 2008; 3: 117-22.

8. Beheshti M, Talebi M, Ardebili A, Bahador A, Lari AR. Detection of AdeABC efflux pump genes in tetracycline-resistant Acinetobacter baumannii isolates from burn and ventilator-associated pneumonia patients. J Pharm Bioallied Sci. 2014; 6: 229-32.

9. Li H, Wang X, Zhang Y, Zhao C, Chen H, Jiang S, et al. The role of RND efflux pump and global regulators in Tigecycline resistance in clinical Acinetobacter baumannii isolates. Future Microbiol. 2015; 10: 337-46.

10. Savari M, Ekrami A, Shoja S, Bahador A. Plasmid borne Carbapenem-Hydrolyzing Class D β -Lactamases (CHDLs) and AdeABC

efflux pump conferring carbapenem-Tigecycline resistance among Acinetobacter baumannii isolates harboring TnAbaRs. Microb Pathog. 2017; 104: 310-17.

11. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; Approved standard. Document M100-S120, 20th ed, Wayne, 2013; PA, USA.

12. Peleg AY, Adams J, Paterson DL. Tigecycline Efflux as a Mechanism for Nonsusceptibility in Acinetobacter baumannii. Antimicrob Agents Chemother. 2007; 51: 2065-69.

13. Fernando DM, Xu W, Loewen PC, Zhanel GG, Kumar A. Triclosan can select for an AdeIJK-overexpressing mutant of Acinetobacter baumannii ATCC 17978 that displays reduced susceptibility to multiple antibiotics. Antimicrob Agents Chemother. 2014; 58: 6424-31.

14. Azimi L, Namvar AE, Jamali S, Lari AR, Bijari A, Lari AR. Relative Expression of Efflux Pumps in Multi Drug Resistant Pseudomonas Aeruginosa. Roum Arch Microbiol Immunol. 2015; 74: 86-90.

15. Pfaffl MW, Lange IG, Daxenberger A, Meyer HH. Tissue-specific expression pattern of estrogen receptors (ER): quantification of ER alpha and ER beta mRNA with real-time RT-PCR. APMIS. 2001; 109: 345-55.

16. Azimi L, Ebrahimzadeh Namvar A, Rastegar Lari A, Jamali S, Rastegar Lari. Comparison of Efflux Pump Involvement in Antibiotic Resistance among Pseudomonas aeruginosa Isolates of Burn and Non-Burn Patients. Arch Pediatr Infect Dis. 2016; 4: e36160.

17. Bijari A, Azimi L, Fallah F, Ardebili A, Lari ER, Lari AR. Involvement of the Multidrug Efflux Pumps in Betalactams Resistant Pseudomonas aerugionsa Clinical Isolates Collected from Burn Patients in Iran. Infect Disord Drug Targets. 2016; 16:172-77.

18. Peleg AY, Adams J, Paterson DL. Tigecycline Efflux as a Mechanism for Nonsusceptibility in Acinetobacter baumannii. Antimicrob Agents Chemother. 2007; 51: 2065-69. 19. Deng M, Zhu MH, Li JJ, Bi S, Sheng ZK, Hu FS, et al. Molecular Epidemiology and Mechanisms of Tigecycline Resistance in Clinical Isolates of Acinetobacter baumannii from a Chinese University Hospital. Antimicrob Agents Chemother. 2014; 58: 297-303.

20. Yuhan Y, Ziyun Y, Yongbo Z, Fuqiang L, Qinghua Z. Over expression of AdeABC and AcrAB-TolC efflux systems confers Tigecycline resistance in clinical isolates of Acinetobacter baumannii and Klebsiella pneumoniae. Rev Soc Bras Med Trop. 2016; 49: 165-71. 21. Yang YS, Chen HY, Hsu WJ, Chou YC, Perng CL, Shang HS, et al. Overexpression of AdeABC efflux pump associated with tigecycline resistance in clinical Acinetobacter nosocomialis isolates. Clin Microbiol Infect. 2018 Jun 12. pii: S1198-743X(18)30473-7.

22. Hu C, Li Y, Zhao Z, Wei S, Zhao Z, Chen H, Wu P. In vitro synergistic effect of amlodipine and imipenem on the expression of the AdeABC efflux pump in multidrugresistant Acinetobacter baumannii. PLoS One. 2018; 1: 13(6):e0198061.